Re-appraisal of the Tocopheroxyl Radical Reaction with $\hat{\beta}$ -Carotene: Evidence for Oxidation of Vitamin E by the β -Carotene Radical Cation

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Accepted by Prof. H. Sies

(Received 23 June 1997; In revised form *11 September 1997)*

Photobleached β -carotene (Car) is regenerated in hexane on a microsecond timescale in the presence of α -tocopherol (TOH) but not when α -tocopherol is absent, as studied by laser flash photolysis. β -Carotene radical cations (Car⁺) likewise react with (excess) α -tocopherol: Car⁺ + TOH \rightarrow Car + TO· + H⁺ (secondorder rate constant of $k = 1.7 \pm 0.1 \cdot 10^7$ M⁻¹ s⁻¹ in homogeneous di-tert-butylperoxide / benzene at 20°C) rather than α -tocopheroxyl radicals (TO \cdot) reacting with β -carotene. In hexane, hexane radicals formed by pulse radiolysis react considerably faster with β-carotene $(k = 2.1 \pm 0.1 \cdot 10^9 \text{ M}^{-1} \text{ s}^{-1})$ than with α -tocopherol $(k = 4.9 \pm 0.1 \cdot 10^6 \text{ M}^{-1} \text{ s}^{-1})$. No evidence was obtained for a slower rate of β -carotene radical cation formation in β -carotene/ α -tocopherol mixtures resulting from α -tocopheroxyl radical oxidation of β -carotene. Steady-state radiolysis experiments confirmed that α -tocopherol protects β -carotene from oxidation by hexane radicals. In both solvent systems, β-carotene is regenerated from the radical cation by α -tocopherol r ather than α -tocopherol being regenerated by β -carotene from the a-tocopheroxyl radical.

Keywords: @-carotene, a-tocopherol, p-carotene radical cation, a-tocopheroxyl radical, laser flash photolysis, pulse radiolysis

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INTRODUCTION

 α -Tocopherol (TOH) is the most important lipidsoluble radical scavenging antioxidant in cellular membranes, plasma and lipoprotein.^[1,2] Other lipid-soluble antioxidants including the ubiquinones and carotenoids are also considered potent free radical scavengers capable of preventing the chain of lipid peroxidation.^[3-6] In both cases, their antioxidant activities may be due to a direct free radical scavenging mechanism or an indirect mechanism which involves re-cycling of α -tocopherol. Free radical scavenging by α -tocopherol involves the formation of the α -tocopheroxyl radical (TO·). Radical interactions with the carotenoids is less understood but generally involves either electron transfer and radical cation (Car⁺) formation or radical-addition to generate carotenoid adduct-radicals.^[7-11] α -Tocopherol and β -carotene exert a synergistic

effect in inhibiting radical-initiated lipid peroxidation in membranes. a-Tocopherol was found to protect β -carotene from oxidation but the mechanisms involved remain a matter for debate.^[12] Carotenoids have recently been shown to reduce the phenoxyl radical to phenol, and it was speculated whether carotenoids had a function in biological systems to recycle phenolic antioxidants by one-electron transfer forming the carotenoid radical cation.^[13,14] The regeneration of phenolic antioxidants by carotenoids may also involve ascorbic acid as the final electron donor reacting with the carotenoid radical cation $[15]$ to export potentially damaging free radical oxidants from lipid-rich environments. The relative reactivity of phenoxylic radicals is strongly dependent on the substitution pattern of the phenol. In another recent study, it was thus demonstrated that phenoxyl radicals generated from phenol by peroxidase was able to oxidize β -carotene whereas the phenoxyl radical of **2,2,5,7,8-pentamethyl-6-hydroxychromane** (a tocopherol homologue) could not,^[16] thereby confirming the lower reactivity of the latter. The delicate balance in the antioxidant hierarchy was nicely demonstrated in the interaction between lycopene and the tocopherol homologues.^[17] a-Tocopherol was thus found to reduce the lycopene radical cation whereas lycopene was able to reduce the δ -tocopheroxyl radical. The relative reactivity of the β - and γ -tocopheroxyl radicals was comparable to that of the lycopene radical cation, and a free radical equilibrium was demonstrated between lycopene and β - and y-tocopherol. It has recently been claimed in a pulse radiolysis study that the α -tocopheroxyl radical could be reduced by most carotenoids including β -carotene but not astaxanthin^[18] via reaction **(1).**

$$
TO \cdot + Car + H^+ \rightarrow TOH + Car^{*+}.
$$
 (1)

All the second-order rate constants were close to the diffusion controlled limit $(5.3 - 26 \cdot 10^9 \,\mathrm{M}^{-1} \,\mathrm{s}^{-1})$.

In light of studies which have provided little evidence supporting reaction **(1)** in model systems, we carried out experiments combining laser flash photolysis and pulse radiolysis utilising three different solvent systems to establish the validity of reaction (1). No evidence supporting reaction (1) was obtained, in fact, the reverse reaction involving the oxidation of α -tocopherol by the β-carotene radical cation was shown to occur.

MATERIALS AND METHODS

Materials

0-Carotene was supplied by Roche A/S (Hvidovre, Denmark) sealed in an ampoule under argon and was used without further purification. Phenol (p.a.) and benzene (p.a.) from Merck (Darmstadt, Germany), hexane (Certified) from Fisher Scientific UK Ltd. (Loughborough, UK) and di-tert-butylperoxide from Merck-Schuchardt (Hohenbrunn bei Miinchen, Germany) were **all used** as received. Anhydrous hexane and vitamin E [2,5,7,8-tetramethyl-2-(4',8', **12'-trimethyltridecyl)-6-chromanol,** a-tocopherol] were obtained from Aldrich (Poole, UK). Nitrous oxide and nitrogen gases were obtained from the British Oxygen Company (Gillingham, UK).

Laser Flash Photolysis

Laser flash photolysis experiments were carried out with an LKS.50 laser flash photolysis spectrometer from Applied Photophysics Ltd (Leatherhead, UK). The third harmonic at 355 nm (50 mJ per pulse) or the fourth harmonic at *266* nm (80 mJ per pulse) of a pulsed Q-switched Nd-YAG laser, Spectron Laser Systems (Rugby, UK), was used for excitation. A 1P28 photomultiplier tube from Hamamatsu (Hamamatsu City, Japan) was used to detect transient absorption changes in the visible region. Red and near infrared detec-

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tion was conducted with an S1336-44BK silicon photodiode from Hamamatsu (Hamamatsu City, Japan). For red and near infrared measurements, red bandpass filters were used in order to minimize degradation of β -carotene by the Xe arc lamp used for monitoring, whereas a *UV* cut-off filter was used for monitoring in the blue-green spectral region. Spectral slit widths were 4-5 nm. The samples were excited in $1 \text{ cm} \times 1 \text{ cm}$ fluorescence cells from Hellma (Miillheim, Germany). All samples were thermostated at 20.0 ± 0.5 °C.

Solutions contained 10 uM B-carotene and α -tocopherol in the mM range. Two sets of solutions were made; one containing 1.75 M phenol in a mixture of di-tert-butylperoxide and benzene (70:30, v/v) with β -carotene and α -tocopherol, and another of β -carotene and α -tocopherol in hexane.

Pulse Radiolysis

The pulse radiolysis facility at the Gray Laboratory Cancer Research Trust has been described previously.^[19] In this study 500 ns pulses of 6 MeV electrons were used to deliver doses of *ca.* 9 Gy as determined by thiocyanate dosimetry.[20] Near infrared detection of carotenoid radicals was performed using a tungsten lamp and a Si photodiode. Bleaching of β -carotene and formation of the α -tocopheroxyl radical in the blue-green spectral region was monitored by photomultiplier utilising a Xe lamp. Two solvent systems were used including an N₂O-saturated hexane system and an $N₂$ -saturated tert-butanol/water (60:40%) system containing 20 mM β -mercaptoethanol.

Steady-State y-Radiolysis

Steady-state radiolysis experiments were performed in a ^{60}Co y-source with nominal activity of 2000 Ci. Solutions were irradiated in gas-tight vials at a rate of 6 Gy min^{-1} as determined by Fricke dosimetry.^[21] The degradation of β -carotene was monitored at 450 nm using a Hewlett Packard 8452A Diode Array Spectrophotometer. The radiation chemical yield for the loss of β-carotene, -G (β-Carotene)/ $μ$ mol J⁻¹ were calculated from the slopes of the linear plots of β -carotene concentration versus radiation dose in Gy.

RESULTS

Laser Flash Photolysis

Laser flash photolysis at 355 nm of di-tertbutylperoxide results in homolytic fission via reaction *(2)*

$$
(CH3)3 COOC(CH3) + h\nu \rightarrow 2(CH3)3 CO: (2)
$$

Quenching of the resultant alkoxyl radical with a large excess of phenol occurs within 10 ns to generate the phenoxyl radical:

$$
(\text{CH}_3)_3\text{CO} + \text{C}_6\text{H}_5\text{OH}
$$

$$
\rightarrow (\text{CH}_3)_3\text{COH} + \text{C}_6\text{H}_5\text{O}. \quad (3)
$$

The phenoxyl radical then abstracts an electron from β -carotene to give the β -carotene radical cation^[13,14] or abstracts a hydrogen atom from α -tocopherol to generate the α -tocopheroxyl radical^[22] via reactions (4) and (5) respectively:

$$
C_6H_5O \cdot + Car \rightarrow C_6H_5O^{-} + Car^{\star+}
$$
 (4)

$$
C_6H_5O^{\bullet} + TOH \rightarrow C_6H_5OH + TO^{\bullet}.
$$
 (5)

The phenoxyl radical reacts with β -carotene^[13] and α -tocopherol^[22] at comparable rates with k_4 = $2 \cdot 10^9$ M⁻¹ s⁻¹ and $k_5 = 1 \cdot 10^9$ M⁻¹ s⁻¹, respectively. In the absence of α -tocopherol, the β -carotene radical cation is rapidly formed (Fig. **1A)** and decays bimolecularly on a subsecond time scale^[13] according to reaction (6):

$$
2\text{Car}^{++} \rightarrow \text{Products(s)}.\tag{6}
$$

FIGURE 1 Time traces of transient absorption following laser flash photolysis (8 ns) at 355 nm of (a) 10 μM β-carotene, 1 mM α-tocopherol and 1.75 M phenol and (b) 10 μM β-carotene and 1.75 M phenol and (b) 10 μM β-carote (p-carotene radical cation absorption) and **(B)** 480 nm (p-carotene absorption). The insert in (A) shows the observed pseudo firstorder rate constant of reaction (7) as a function of α -tocopherol concentration.

In the presence of a large excess of α -tocopherol (100-fold), most phenoxyl radicals are scavenged via reaction (5) and only a small amount of p-carotene radical cation is formed via reaction **(4)** (Fig. 1A). However, the lifetime of the β -carotene radical cation is much shorter in the presence of α -tocopherol, and disappears completely in less **than 100 μs. This indicates that the β-carotene rad-** ical cation is scavenged by α -tocopherol to generate the α -tocopheroxyl radical via reaction (7):

$$
Car^{++} + TOH \rightarrow Car + TO^+ + H^+. \tag{7}
$$

The decay was exponential and first-order in α -tocopherol concentration. By varying the concentration of a-tocopherol, the rate constant of

reaction (7) was determined to be $k_7 = 1.7 \pm 0.1$ \cdot 10^7 M⁻¹ s⁻¹ from the linear slope of the observed rate constant versus α -tocopherol concentration in Fig. **1A.** Kinetic traces recorded in the 400-500 nm spectral region (where both the β -carotene ground-state absorption and the a-tocopheroxyl radical absorption occur) confirmed reaction **(7).** In the absence of α -tocopherol, a positive transient is observed at short times at 480 nm (Fig. **1B)** due to the formation of the phenoxyl radical (reaction (3)), and bleaching of β -carotene via reaction **(4)** is observed on a longer time scale. In the presence of α -tocopherol, no bleaching of p-carotene is observed but only formation of the α -tocopheroxyl radical which has a very long lifetime in this system.^[22] This is consistent with the kinetic observations of the β -carotene radical cation (Fig. **1A)** and is entirely consistent with reaction (7). The complete inhibition of β -carotene degradation by the phenoxyl radical in the presence of α -tocopherol indicates that the resultant a-tocopheroxyl radicals generated in reaction (5) do not react with β -carotene via reaction (1) as has been suggested.[18]

The α -tocopheroxyl radical may also be generated directly by photolysis at 266 nm in hexane

$$
TOH + h\nu \rightarrow TO+H.
$$
 (8)

The hydrogen atom is rapidly $(k = 1.5$ 10^8 M⁻¹ s⁻¹)^[23] scavenged by hexane

$$
H^{\bullet} + C_6 H_{14} \rightarrow H_2 + C_6 H_{13}. \tag{9}
$$

The α -tocopheroxyl radical generated by photolysis has a rather short lifetime (Fig. *2A)* compared to the lifetime in di-tert-butylperoxide / benzene (Fig. 1B). The lifetime, however, is greatly increased in the presence of β -carotene (Fig. 2A). The short lifetime may be explained by the reaction between the a-tocopheroxyl radical and the hexane radical :

$$
TO \cdot + C_6H_{13} \cdot \rightarrow \text{product(s)}.
$$
 (10)

FIGURE 2 Time traces of transient absorption following laser flash photolysis (8 ns) at 266 nm of **(a)** 1 mM a-tocopherol, **(b)** 10 μ M β -carotene and **(c)** 10 μ M β -carotene and 1 mM α -tocopheroxyl radical absorption), (B) 450 nm (β -carotene absorption) and (C) 1040 nm (β -carotene radical cation absorption).

In the presence of β -carotene the lifetime of the a-tocopheroxyl radical is increased because the hexane radical is scavenged by β -carotene in reaction (11):

$$
Car + C6H13 \rightarrow Car+ + C6H13.
$$
 (11)

The increased lifetime of the α -tocopheroxyl radical in the presence of β -carotene indicates that β -carotene effectively competes with the α -tocopheroxyl radical for the hexane radical. Despite the much higher concentration of *a*tocopherol than of the α -tocopheroxyl radical and β -carotene, α -tocopherol does not seem to compete effectively for the hexane radical

$$
TOH + C_6H_{13} \rightarrow TO + C_6H_{14}
$$
. (12)

If it did, a much longer lifetime of the α tocopheroxyl radical were to be expected, and besides the instantaneous formation of α -tocopheroxyl radical due to photolysis, a slower formation due to reaction 12 would be expected. Neither is observed (Fig. **2A).** The decay kinetics of the α -tocopheroxyl radical does not change in the presence of oxygen, indicating that either the hexane radicals react faster with the α tocopheroxyl radical than with oxygen or the hexylperoxyl radicals formed by reaction between hexane radicals and oxygen react just as fast with the α -tocopheroxyl radical as the hexane radical does. β -Carotene is bleached to a small extent by the laser pulse at **266** nm (Fig. **2B).** In the presence of α -tocopherol the extent of bleaching is higher due to reaction (11). However, p-carotene is only regenerated in the presence of α -tocopherol (via reaction (7)) but not in the absence of a-tocopherol (Fig. **2B).** Fig. 2C shows that the β -carotene radical cation is generated to a larger extent in the presence of α -tocopherol, in accordance with reaction (11). These results provide further support for reaction (7) but once again no evidence was obtained for the reverse reaction **(1).**

Pulse Radiolysis

A previous pulse radiolysis study of a-tocopherol/ carotenoid mixtures in hexane claimed to provide evidence for the oxidation of different carotenoids by the α -tocopheroxyl radical via reaction (1) .^[18] In light of our laser photolysis experiments which provided evidence for reaction **(7)** rather than reaction (l), further pulse radiolysis experiments were performed in N_2O -saturated hexane. Hexane radicals generated by radiolysis react with both β -carotene and α -tocopherol to generate the β -carotene radical cation and the α -tocopheroxyl radical, respectively. The relative reactivity of hexane radicals toward the two antioxidants is significantly different. Pulse radiolysis of α -tocopherol in hexane generated transient absorption with a maximum at 420 nm (Fig. **3A)** attributable to the α -tocopheroxyl radical.^[18,24] The formation of the α -tocopheroxyl radical was exponential and first-order in (excess) α -tocopherol concentration (0.1-8 mM). Plotting the observed first-order rate constant against a-tocopherol concentration vielded the second-order rate constant $k = 4.9 \pm 0.1$ \cdot 10⁶ M⁻¹ s⁻¹ (insert Fig. 3A). The lifetime of the a-tocopheroxyl radical generated by pulse radiolysis is much longer than the lifetime when generated by photolysis (compare Fig. **3A** with Fig. *2A)* probably because of reaction **(10)** in the latter. The formation of the β -carotene radical cation at 999 nm^[18,25] was also exponential and first-order in β -carotene concentration (10-80 μ M) as shown in Fig. **3B** but occurs significantly faster at a rate close to the diffusion controlled limit $k = 2.1 \pm 0.1 \cdot 10^9$ M⁻¹ s⁻¹. In marked contrast to the α -tocopheroxyl radical, the β -carotene radical cation is relatively short-lived in hexane and decays bi-molecularly to products via reaction **(6)** in less than 200 µs at the same dose per pulse *ca*. 3 Gy. Lower doses per pulse could not be used since the radiation chemical yield of the radical cation in hexane is very low (a fairly large dose per pulse of **3** Gy resulted in radical cation absorption as low as *cu.* 1 milli-absorbance unit at **999 nm).**

It has been previously reported^[18] that the rate of p-carotene radical cation formation in a 10 **pM** β -carotene/100 µM α -tocopherol mixture is much slower than with 10 μ M β -carotene alone. This slower formation of the radical cation has been ascribed to reaction **(1)** where electron transfer occurs from the carotenoid to the α tocopheroxyl radical.^[18] We obtained no evidence for a slower rate of radical cation formation in the presence of 100 μ M α -tocopherol. Indeed, Fig. **4B** clearly shows that the observed rate of formation of the B-carotene radical cation increased with increasing concentrations of a-tocopherol **(0.1-33** mM) at constant dose per pulse $(ca. 2.9 Gy)$ and constant β -carotene concentration (20 μ M). The increase in rate was also accompanied by an increase in β -carotene radical cation absorption at **999** nm (Fig. **4A).** In order to ascertain whether this was due to reaction **(l),**

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FIGURE 3 (A) Typical transient absorption of the α-tocopheroxyl radical generated by pulse radiolysis (9 Gy) of N₂O-saturated hexane containing 4.1 mM a-tocopherol. The insert in **(A)** shows the observed pseudo first-order rate constant for a-tocopheroxyl radical formation as a function of a-tocopherol concentration. Panel **(B)** shows a typical transient absorption of the p-carotene radical cation generated by pulse radiolysis (2.9 Gy) of N₂O-saturated hexane containing $20 \mu M$ β -carotene. The insert in panel (B) shows the observed pseudo first-order rate constant for β -carotene radical cation formation as a function of β -carotene concentration.

FIGURE 4 In panel A, pulse radiolysis (2.9 Gy) of 20 μ M β -carotene in N₂O-saturated hexane generates the β -carotene radical cation **(a).** The p-carotene radical cation absorption increases on the introduction of **(b)** 200 **pM** and **(c) 33** mM a-tocopherol. Panel B shows the increase in the observed pseudo first-order rate constant for β -carotene radical cation formation as a function of α -tocopherol concentration at constant dose (2.9 Gy) and β -carotene concentration (20 μ M).

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further experiments were performed in *tert*butanol/water mixtures containing 20 mM ßmercaptoethanol and various β -carotene and a-tocopherol concentrations. The thiyl radicals **(RS.)** generated in this system react exclusively with β -carotene to generate adduct-radicals **[RS-Car].** in reaction **(13),** and unlike many other radical species thiyl radicals are unique in that they do not produce the β -carotene radical cation.^[7,19] Thiyl radicals also abstract a hydrogen atom from α -tocopherol to generate the a-tocopheroxyl radical according to reaction **(14).**

RS⁺
$$
\text{Car} \rightarrow [\text{RS} - \text{Car}]
$$
 (13)

$$
RS \cdot + TOH \rightarrow RSH + TO \cdot (14)
$$

Thus, any β -carotene radical cation formation in this system could **only** be attributable to reaction **(1).** Pulse radiolysis of excess a-tocopherol (-10 mM) in the presence of $20 \mu \text{M}$ β -carotene did not generate any radical cation absorption despite the fact that the α -tocopheroxyl radical is stable for ms after the electron pulse.

In the hexane system the decay of the **B-carotene** radical cation changed from pure second-order kinetics (Fig. 5, transient a) to first-order in the presence of excess α -tocopherol (Fig. 5, transient b). The $mixed-order$ kinetics associated with β -carotene

FIGURE 5 The decay of the @-carotene radical cation in the absence of a-tocopherol **is** second-order **(a)** and exhibits mixed-order kinetics in the presence of **33** mM a-tocopherol **(b).** The insert shows the increase in the observed pseudo first-order rate constant for β -carotene radical cation decay with an increase in α -tocopherol concentration at constant dose (2.9 Gy) and β -carotene concentration (20 μ M).

radical cation decay was resolved into the contributing first-order component due to reaction (7) and the second-order component of the competing reaction (6). The slope of the linear plot of observed first-order rate constant versus α -tocopherol concentration gave the rate constant $k_7 = 2.0 \pm 0.1 \cdot 10^4$ M^{-1} s⁻¹ which is approximately three orders of magnitude slower than that determined by laser photolysis in the di-tert-butylperoxide / benzene system. Though we have no ready explanation for this large difference in rate constant between this system and the phenol/ di-terf-butylperoxide / benzene system, the difference could be due to differences in reaction medium.

Further evidence supporting these observations was derived from simple steady-state radiolysis experiments of β -carotene/ α -tocopherol mixtures in $N₂O$ -saturated hexane. Under identical experimental conditions, the radiation chemical yields for the loss of the antioxidants are $-G(\beta$ -Carotene) = 0.15 µmol J⁻¹ and $-G(\alpha$ tocopherol) = $0.21 \mu \text{mol}$ J⁻¹, respectively. Pulse radiolysis and laser photolysis experiments have demonstrated the stability of the α -tocopheroxyl radical in hexane. If oxidation of β -carotene by the α -tocopheroxyl radical via reaction (1) was taking place in this system, then the irradiation of β -carotene in the presence of excess α -tocopherol should result in a *ca*. 29% increase in β-carotene degradation. Figure **6** shows a plot of -G(P-Carotene) / μ mol J⁻¹ *versus* α -tocopherol concentration (0-500 μ M) which clearly demonstrates that excess α -tocopherol (500 μ M) completely suppresses hexane radical-induced degradation of β-carotene (80 μ M). If reaction (1) could occur in this system, then the complete scavenging of hexane radicals by α -tocopherol should not have offered any protection. The reactivity of hexane radicals toward β -carotene is greater than with a-tocopherol by over three orders of magnitude (see Fig. **3). As** shown in Figure 6 when [α -tocopherol] < [β -carotene], the hexane radicals are primarily scavenged by β -carotene but α tocopherol still offers protection. **A** low dose rate of 6.2 Gy min^{-1} was used to minimise radicalradical reactions, mainly the bimolecular decay of the β -carotene radical cation via reaction (6). The mechanism of protection may therefore be due to the scavenging of the β -carotene radical cation by a-tocopherol and regeneration of β -carotene via reaction (7).

DISCUSSION

In the recent paper by Böhm and colleagues $[18]$ a slower formation of the β -carotene radical cation in the presence of 100 μ M α -tocopherol than in the absence of α -tocopherol was observed upon pulse radiolysis in hexane. This slower formation was ascribed to reaction between the α -tocopheroxyl radical and β -carotene. The rate of this reaction was found to be close to diffusion controlled for a number of carotenoids (in the case of zeaxanthin even higher than diffusion controlled). The only exception was astaxanthin where evidence was obtained for the reverse reaction (7) where the carotenoid radical cation oxidised α -tocopherol to the α -tocopheroxyl radical. Our laser flash photolysis and radiolysis experiments clearly demonstrate that reaction (7) appears to be the rule rather than the exception (at least in the case of β -carotene).

Pulse radiolysis of β -carotene/ α -tocopherol mixtures in hexane is an unnecessarily complicated system for the study of the interaction of these *two* antioxidants. The increase in β-carotene radical cation absorption in the presence of increasing concentrations of α -tocopherol may possibly be due to another α -tocopheroxyl radical species such as the α -tocopherol radical cation (TOH $^{\text{+}}$) which may generate the β -carotene radical cation following electron transfer from β carotene

$$
TOH^{+} + Car \rightarrow Car^{+} + TOH.
$$
 (15)

Reaction (15) may also explain the observed increase in the rate of β -carotene radical cation formation in the presence of α -tocopherol. Generally,

FIGURE 6 Variation of the radiation chemical yield of β -carotene degradation, -G(β -Carotene)/ μ mol J⁻¹ during the steady-state γ -radiolysis of 80 µM β -carotene in N₂O-saturated hexane with increasing α -tocopherol concentration. Excess α -tocopherol completely inhibits β -carotene degradation. The insert shows the corresponding degradation of the β -carotene as a function of radiation dose in the presence of various concentrations of α -tocopherol.

phenoxyl radical cations have pK_a values of $ca. 5.$ ^[26] The α -tocopherol radical cation would be expected to deprotonate very rapidly at neutral pH in aqueous systems but is very stable in acidic acetonitrile where the rate of deprotonation to the α -tocopheroxyl radical is considerably slower.^[27] In the present study no spectral evidence for a tocopherol radical cation was obtained in hexane so the importance of this species in the hexane system remains a matter for debate.

Generating the α -tocopheroxyl radical by laser flash photolysis in the presence of β -carotene helped to circumvent competition kinetics associated between reactions **(6)** and **(7)** in the hexane system. Despite the fact that the rate constant $k_7 = 1.7 \cdot 10^7$ M⁻¹ s⁻¹ has been determined in homogeneous solution it is sufficiently fast to at least implicate reaction (7) in carotenoid/ α tocopherol interactions within heterogeneous lipophilic environments particularly cell membranes and lipoprotein. Based on this investigation we have found no evidence that β -carotene can recycle the a-tocopheroxyl radical back to *a*tocopherol. However, our results do indicate that α -tocopherol recycles β -carotene by scavenging the β -carotene radical cation.

CONCLUSIONS

Previous studies of the interaction between *a*tocopherol and carotenoids have indicated that the α -tocopheroxyl radical cannot be reduced by carotenoids but that the carotenoid radical cation may be reduced by α -tocopherol.^[12,16,17] The present results have clearly demonstrated that this is indeed the case, and a rate constant $k_7 = 1.7 \pm 0.1$ 10^7 M⁻¹ s⁻¹ for this reaction has been determined. Our results are consistent with the hypothesis that α -tocopherol protects β -carotene during membrane-associated oxidative stress and not *vice versa.*^[18]

Acknowledgements

The authors thank Professor Catherine Rice-Evans (International Antioxidant Research Group, UMDS-Guy's Hospital, London) for helpful discussions. Roche A/S is kindly thanked for providing the β -carotene. This work is funded by the F0TEK programme through LMC-Center for Advanced Food Studies. **Dr.** Steven A. Everett is supported by the Cancer Research Campaign [CRC] and Dr. Amadeus Willnow by the Association for International Cancer Research [AICR].

Note added in proof

Recent findings by Val-gimigli *et* a1.[281 corroborate our findings, i.e. that β -carotene cannot reduce the α -tocopheroxyl radical.

References

- Burton G. W., Joyce A. and Ingold K. **U. (1982).** First proof that vitamin **E** is major lipid-soluble, chain-breaking antioxidant in human blood plasma. *Lancet,* **2,327.**
- Burton *G.* W., Joyce **A.** and Ingold K. U. **(1983).** Is vitamin E the only lipid-soluble, chain-breaking antioxidant in human blood plasma and erythrocyte membranes? *Archives* of *Biochemistry and Biophysics,* **221, 281-290.**
- **[3]** Ingold K. **U.,** Bowry V. W., Stocker R. and Walling C. **(1993).** Autoxidation of lipids and antioxidation by atocopherol and ubiquinol in homogeneous solution and in aqueous dispersions **of** lipids: Unrecognized consequences of lipid particle size as exemplified by oxidation of human low density lipoprotein. *Proceedings of the National Academy* of *Sciences,* **90,4549.**
- **[4]** Stocker R., Bowry **V.** W. and Frei B. **(1991).** Ubiquinol-10 protects human low density lipoprotein more efficiently against lipid peroxidation than does a-tocopherol. *Proceedings* of *the National Academy* of *Sciences,* **88, 1646-1650**
- **[5]** Suarna C., Hood R. L., Dean R. T. and Stocker R. **(1993).** Comparative antioxidant activity of tocotrienols and other natural lipid-soluble antioxidants in a homogeneous system, and in rat and human lipoproteins. *Biochimica et Biuphysica Acta,* **1166, 163-170.**
- *[6]* Rice-Evans C. A., Sampson J., Bramley P. M. and Holloway D. E. **(1997).** Why do we expect carotenoids to be antioxidants *in vim? Free Radical Research,* **26,381-398.**
- **[7]** Everett S. A,, Dennis M. F., Pate1 K. B., Maddix S., Kundu S. C. and Willson R. L. **(1996).** Scavenging of nitrogen dioxide, thiyl, and sulfonyl free radicals by the nutritional antioxidant p-carotene. *Journal* of *Biological Chemistry,* **271,3988-3994.**
- **(81** Hill T. J., Land E. J., McGarvey D. J., Schalch W., Tinkler J. H. and Truscott T. G. **(1995).** Interactions between carotenoids and the CC13OOy radical. *Journal of the* American Chemical Society, 117, 8322-8326
- **[9]** Liebler D. C. and McClure T. D. **(1996).** Antioxidant reactions **of** p-carotene: identification of carotenoid-radical adducts. *Chemical Research in Toxicology,* **9,8-11.**
- **[lo]** Mortensen **A.** and Skibsted L. H. **(1996).** Kinetics of photobleaching of p-carotene in chloroform and formation of transient carotenoid species absorbing in the near infrared. *Free Radical Research,* **25,355-368.**
- [ll] Willson **R.** L. **(1983).** Free radical protection: why vitamin **E,** not vitamin C, p-carotene or glutathione? In *Biology* of *vitamin E* (Eds. R. Porter and J. Whelan),
- Pitman Press, London, pp. **19-37. [12]** Palozza P. and Krinsky N. I. **(1992).** p-Carotene and atocopherol are synergistic antioxidants. *Archives* of *Biochemistry and Biophysics,* **297,184-187.**
- **(131** Mortensen A. and Skibsted L. H. **(1996).** Kinetics of parallel electron transfer from β -carotene to phenoxyl radical and adduct formation between phenoxyl radical and p-carotene. *Free Radical Research,* **25,515-523.**
- **[14]** Mortensen A. and Skibsted L. H. **(1997).** Importance of carotenoid structure in radical-scavenging reactions. *Journal* of *Agricultural and Food Chemistry,* **95,2970-2977.**
- **[15]** Truscott T. G. **(1996).** p-Carotene and disease: a suggested pro-oxidant and anti-oxidant mechanism and speculations concerning its role in cigarette smoking. *Journal* of *Photochemistry and Photobiology B: Biology,* **35, 233-235.**
- **(161** Tyurin V. **A.,** Carta G., Tyurina Y. Y ., Banni S., Day B. W., Corongiu F. P. and Kagan V. E. **(1997).** Peroxidasecatalysed oxidation **of** p-carotene in HL-60 cells and in model systems : involvement **of** phenoxyl radicals. *Lipids,* **32,131-142.**
- **[17]** Mortensen A. and Skibsted L. H. **(1997).** Real time detection of reactions between radicals of lycopene and tocopherol homologues. *Free Radical Research* **27,229-234.**
- **(181** Bohm F., Edge R., Land E. J., McGarvey D. J. and Truscott T. G. **(1997).** Carotenoids enhance vitamin **E**

antioxidant efficiency. *Journal* of *the American Chemical Society,* **119,621-622.**

- **[19]** Everett **S. A.,** Kundu *S.* C., Maddix S. and Willson R. L. **(1995).** Mechanisms of free-radical scavenging by the nutritional antioxidant p-carotene. *Biochemical Society Transactions,* **23,230s.**
- **[20]** Bielski 8. **H.** J. **(1993). A** pulse radiolysis study of the reaction of ozone with C12-in aqueous soluhon. *Radiation Physics and Chemisfy,* **41,527-530.**
- **[21]** Tabata **Y.,** It0 Y. and Tagawa **S. (1991).** *Handbook* of *Rudiation Chemistry.* CRC Press, Boca Raton, Florida.
- **[22]** Foti M., Ingold K. **U.** and Lusztyk J. **(1994).** The **surpris**ingly high reactivity of phenoxyl radicals. *Journal* of *the American Chemical Society,* **116,9440-9447.**
- **[23]** Neta **P.,** Fessenden **R.** W. **and** Schuler R. H. **(1971).** An electron spin resonance study of the rate constants for reaction **of** hydrogen atoms with organic compounds in aqueous solution. *Journal* of *Physical Chemistry,* **75, 1654-1666.**
- **[24]** Thomas M. J. and Bielski 8. H. J. **(1989).** Oxidation and reaction of trolox c, a tocopherol analogue, in aqueous solution. A pulse radiolysis study. *Journal* of *the American Chemical Society,* **111, 3315-3319.**
- [25] Dawe E. A. and Land E. J. **(1975).** Radical ions derived from photosynthetic polyenes. *Journal* of *the Chemical Society Faraday Transactions* **1,71,2162-2169.**
- **[26]** Land E., Porter G. and Strachan E. **(1960).** Primary photochemical processes in aromatic molecules. *Transactions* of *the Faraday Society,* **57,18851893.**
- 1271 Svanholm U., Bechgaard K. and Parker V. D. **(1974).** Electrochemistry in media of intermediate acidity. VIII. Reversible oxidation products of the α -tocopherol model compound. Cation radical, cation, and dication. *Journal* of *the American Chemical Society,* **96,2409-2413.**
- **[28]** Valgimigli L., Lucarini M, Pedulli G. F. and Ingold K. U. **(1997).** Does p-carotene really protect vitamin **E** from oxidation? *Journal* of *the American Chemical Society,* **119, 80958096.**